

## Localization of *Sry* gene on Y chromosome of *Muntjac munticus vaginalis* \*

ZHANG Yue (张悦)<sup>1</sup>, SHAN Xiangnian (单祥年)<sup>1\*\*</sup>, LIU Ningsheng (刘宁生)<sup>1</sup>,  
LU Xiaoxuan (鲁晓萱)<sup>1</sup>, NIE Wenhui (聂文惠)<sup>2</sup>, YANG Fengtang (杨凤镪)<sup>2</sup>,  
WANG Jinhuan (王金焕)<sup>2</sup> and CHEN Yuze (陈玉泽)<sup>2</sup>

1 Medical Genetics Research Center, Medical College, Southeast University, Nanjing 210009, China

2 Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming 650223, China

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**Abstract** The chromosomes 1,  $Y_1$ ,  $Y_2$  of *Muntjac munticus vaginalis* were isolated by fluorescence activated chromosome sorting and amplified by degenerate oligonucleotide primed-polymerase chain reaction (DOP-PCR). A primer pair within human *Sry* HMG box was designed and the *Sry* gene of the male *M. m vaginalis* was amplified. The product was cloned and sequenced. The result proved that *Sry* is located on chromosome  $Y_2$ , which is the sex-determining chromosome in the male *M. m vaginalis*.

**Keywords:** *M. m vaginalis*, *Sry* gene, DOP-PCR, sex.

The Indian muntjac (*Muntjac munticus vaginalis*) has the lowest diploid chromosome number ( $2n = 6$  in the female, 7 in male) so far discovered in mammals<sup>[1]</sup>. The male *M. m muntjac* has two Y chromosomes. Which one takes part in sex determination or do both of them have the function? This is a puzzling question.

People did not know whether mammalian sex determination depends on the Y chromosome until the late 1950s, and it was proposed that there should be a testis-determining factor on Y chromosome, which determines the genital ridge developing into the testis. In 1990, the SRY gene from sex-determining region of the Y chromosome was isolated in humans by Sinclair et al.<sup>[2]</sup> And later on it was identified as a conserved gene on the Y chromosome of all animals. In 1991, the mouse homologues *Sry* was shown to express in the genital ridge just before testis differentiation, and its identity as the testis-determining factor was irrefutably demonstrated by the male development of XX mice transgenic for a 14kb DNA containing *Sry*<sup>[3]</sup>.

In this study, we used chromosome sorting, degenerate oligonucleotide primed-polymerase chain reaction (DOP-PCR) and amplification of the *Sry* gene of the male *M. m vaginalis* to localize *Sry* gene on the chromosomes 1,  $Y_1$ ,  $Y_2$  of *M. m vaginalis*. The results showed that  $Y_2$  is the sex-determining chromosome in the male *M. m vaginalis*.

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\*\* Corresponding author, E-mail: shanxn@jlonline.com

## 1 Materials and methods

### 1.1 Cells

The cells of male *M. m vaginalis* derived from embryonic lung were provided by Kunming Institute of Zoology, the Chinese Academy of Sciences.

### 1.2 Methods

**1.2.1 Chromosome preparation and sorting.** Harvest and preparation of chromosomes for cytogenetic analyses followed standard procedures. Before harvesting, the cells were treated with colcemid at a final concentration of 0.1  $\mu\text{g}/\text{mL}$ , then hypotonized and centrifugated. The pellet was resuspended in 500  $\mu\text{L}$  PAB buffer, then the suspension was stained by the addition of 40  $\mu\text{g}/\text{mL}$  chromomycin A3, 2mmol/L  $\text{MgSO}_4$  and 2  $\mu\text{g}/\text{mL}$  of Hoechst 33258 and left on ice for at least 2h. Fifteen minutes prior to flow analysis,  $\text{Na}_2\text{SO}_4$  and  $\text{C}_6\text{H}_7\text{NaO}_7$  were added to the final concentrations of 10 and 25 mmol/L, respectively. The stained chromosome suspensions were sorted on an FACStar Plus flow sorter (Becton Dickinson), and the chromosomes 1,  $Y_1$ ,  $Y_2$  were isolated<sup>[4]</sup>.

**1.2.2 DOP-PCR.** The primer used was: 5'-CCGACTCGAGNNNNNNATGTGG-3' (in which N = A, C, G, or T in approximately equal proportions). PCR was carried out in 2.5U *Taq* polymerase, 0.2 mmol/L dNTP, 50 pmol primer, 50 ng template in a volume of 100  $\mu\text{L}$  reaction. After an initial denaturation for 9 min at 94  $^{\circ}\text{C}$ , 9 cycles of 94  $^{\circ}\text{C}$  for 1 min, 30  $^{\circ}\text{C}$  for 1.5 min, 72  $^{\circ}\text{C}$  for 3 min proceeded, followed by 30 cycles of 94  $^{\circ}\text{C}$  for 1 min, 62  $^{\circ}\text{C}$  for 1 min, and 72  $^{\circ}\text{C}$  for 3 min. A final extension at 72  $^{\circ}\text{C}$  was increased to 10 min. The size of PCR product was determined by running 3 ~ 5  $\mu\text{L}$  products on a 1% agarose gel.

**1.2.3 Amplifying, cloning and sequencing of *Sry* gene of *M. m vaginalis*.** The primers designed for *Sry* of *M. m vaginalis* were 5'-TGAAGCGACCCATGAACG-3' and 5'-CGACGAGGACGATACTTA-3', which were based on the Panda *Sry* gene sequence<sup>[5]</sup>. PCR cycling was performed on 4  $\mu\text{L}$  DOP-PCR products of chromosome 1,  $Y_1$ ,  $Y_2$  respectively, using 2.5 U *Taq* DNA polymerase per 100  $\mu\text{L}$  reaction volume. The terminal cycling included 35 cycles of 1 min denaturation at 94  $^{\circ}\text{C}$ , 1 min annealing at 58  $^{\circ}\text{C}$  and 1 min elongation at 72  $^{\circ}\text{C}$ . Products of amplification were separated on a 2% agarose gel. A 200bp DNA fragment corresponding to the *Sry* gene was identified as expected, and was excised from the gel. This fragment was then purified and cloned into a PGEM-T plasmid vector<sup>[6]</sup>, and transformed to DH5 $\alpha$ . The recombinant clones were identified by color screening. A positive clone was obtained and sequenced.

## 2 Results and discussion

DOP-PCR amplified uncounted fragments from chromosome 1,  $Y_1$ ,  $Y_2$  of *M. m vaginalis*, which showed a smear on the agarose gel (Fig. 1). The size of products ranged from 200 to 2000 bp.

With the primers designed within human SRY HMG box sequence, the *Sry* gene of the male *M. m vaginalis* was amplified only on chromosome  $Y_2$ , but not on chromosome 1 and  $Y_1$  (Fig. 2). The sequence of *Sry* HMG box is shown in Figure 3.

DOP - PCR allows essentially random amplification of DNA from any source. It is based on the

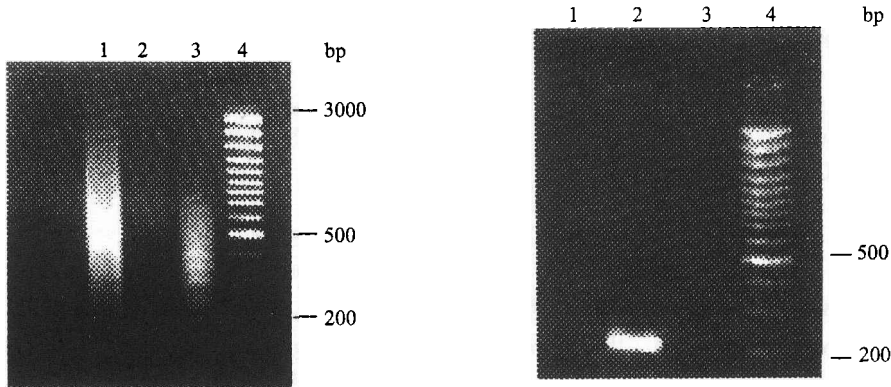


Fig. 1 DOP-PCR products of chromosomes 1,  $Y_1$  and  $Y_2$ . Lane 1,  $Y_1$ ; lane 2,  $Y_2$ ; lane 3, chromosome 1; lane 4, 100 bp DNA marker.

Fig. 2 *Sry* PCR products of chromosomes 1,  $Y_1$  and  $Y_2$ . Lane 1,  $Y_1$ ; lane 2,  $Y_2$ ; lane 3, chromosome 1; lane 4, 100bp DNA marker.

principle that, at a sufficiently low annealing temperature, only 6 specific bases of 3' end of the oligonucleotide will prime the reaction, theoretically priming every  $4^6$  base pairs along the starting DNA. Any specified 6-sequence is therefore likely to occur at a frequency allowing a highly diverse amplification to take place<sup>[7]</sup>. This is an efficient and convenient technique to amplify a small amount of DNA such as single sorted chromosome.

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CCTTCATTGT GTGGTCTCGT GAACGAAGAC GAAAGGTAAC TATAGAGAAT
CCCAAAATGC AAAACTCAGA GATCAGCAAG CAGCTGGGGT ATGAGTGGAA
AAGGCTTACA GATGCTGAAA AGCGCCCAT CTTTGAGGAG GCACAGAGAC
TACTAGCCAT ACACCGAGAC AAATACCCGG GCTAT
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Fig. 3 Sequence of *Sry* HMG box of *Muntjak muntiacus vaginalis*.

Shi et al.<sup>[8]</sup> have conducted many studies on *Muntjacs* at cytogenetic level. They showed that a high degree diversity of karyotype existed in these animals. The chromosome numbers of female and male *Indian muntjac* are different too. Ma et al.<sup>[9]</sup> studied synaptonemal complexes in spermatocytes of Chinese *Muntjac Muntiacus reevesi*, Black *Muntjacu M. crinifrons* and Indian *Muntjac M. Muntjak*. They concluded that the sex-determining karyotype of the Indian *Muntjac* was XAXA/XAY.

We proved that *Sry* gene is located on chromosome  $Y_2$ , and this chromosome should be the real sex-determining chromosome in the male *M. m vaginalis*.

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